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# DEVELOPMENT OF THE APPROACHES FOR COMPLEX UTILIZATION OF BROWN ALGAE (*FUCUS VESICULOSUS*) BIOMASS FOR THE OBTAINING OF VALUE-ADDED PRODUCTS

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The seaweed collected after stranding on beaches of Latvia is underexploited natural resource, which has a potential as raw material for biologically active compound extraction for cosmetic and pharmacy and fertilizer in sustainable agriculture. The aim of the present study was development of the approach for the processing of brown alga *Fucus vesiculosus* biomass, collected from the Gulf of Riga. The thorough characterization of the chemical composition of *Fucus vesiculosus* has shown that it is a potential source for obtaining of nitrogen-containing fertilizers, and biologically active compounds. One of the proposed approaches for the processing of the *Fucus vesiculosus* biomass under study includes algae extraction with organic solvents and CaCl<sub>2</sub> solution and obtaining soil organic amendment on the basis of the extract-free residue. The ethyl acetate extract was rich in phenolic compounds (430 ± 30 GAE mg/g) with high antioxidant activity in DPPH<sup>•</sup> and ABTS<sup>++</sup> tests. The ethanol extract contained significant amounts of phlorotannins that was confirmed by the data of LC-MS/MS analysis. The CaCl<sub>2</sub> extract was used for the obtaining of sulphated polysaccharide fucoidane (yield  $\approx$ 7% on the basis of oven dry matter, o.d.m.), which has numerous biological activities. The extract-free residue didn't show phytotoxicity. The extract free algal biomass, the mechano-chemical treatment of algal biomass with lignin was proposed. EPR analyses confirmed interaction between algal biomass and lignin.

Keywords: Fucus vesiculosus, extracts, antioxidants, organic fertilizers, biorefinery

## **INTRODUCTION**

Latvian bioeconomical development requires effectively utilize available natural resources and seaweeds are one of the underexploited local resources (Bioeconomy strategy..., 2017). The shores of the Baltic Sea in Latvia are almost 500 km long, and thousands tons of seaweeds every year are washed ashore. According to EU Directive 2006/7/EC, washed out algae have to be collected by local municipalities during the beach season in recreation places.

Washed out algae have been used for centuries as a natural fertilizer. However, rapid application of algal biomass is possible not always, but storage of the algal biomass results in emission of odors and release of a liquefied fraction (Michalak *et al.* 2013). Seaweed processing is focused mainly on hydrocolloids production which is large scale and growing industry (Porse *et al.* 2017), however seaweeds treatment plant in Latvia ("Nakotne", Dobele) was closed in early nineties (Grickus *et al.* 2013). One of the reasons for it was extreme decrease of red algae *Furcellaria lumbricalis* habitat in Latvia coastal zone after the oil spill from tanker in 1979 near Ventspils and in 1981 near Klaipeda (Korolev *et al.* 2004). To our knowledge at the present moment "Est-Agar" AS (Estonia) is the only plant in Baltic States which produces the hydrocolloid furcellaran from *Furcellaria lumbricalis*.

The dominant macroalgae in the Baltic Sea and the Gulf of Riga is *Fucus vesiculosus* (Seisuma *et al.* 2011). In many countries brown algae have wide application in several area such as human and animal nutrition, cosmetic and fertilizers (Rioux *et al.* 2015). However, brown algae *F. vesiculosus* can accumulate heavy metals from marine water

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(Seisuma *et al.* 2011) which limits their application for nutrition or medicine. *F. vesiculosus* contain various biologically active compounds with potential market value such as sulfated polysaccharide fuciodan (Ale *et al.* 2011), phlorotannins (Kirke *et al.* 2017) *etc.* The interest in algae based chemicals have increased over the past few years. An effective biorefinery approach for algae processing can only be achieved by integration with other industries. It is necessary to establish a proper connection between the various input and output flows of the products, as well as the services provided by the participating industries. Nowadays many researches are focused on production of biodiesel and biogas from algae (Dong *et al.* 2016, Balina *et al.* 2015). The development of integrated algae biorefinery process capable of producing multiple products is crucial for the commercialization of the Latvian underused valuable resource.

The aim of present study was complex utilization of *Fucus vesiculosus* biomass for the obtaining of high value-added niche products such as natural antioxidants, well known bioactive polisaccharide fucoidan, and fertilizers on the basis of solid residual biomass of *F. vesiculosus* after extraction.

#### MATERIALS AND METHODS

Seaweeds were manually collected on the beach area of Gulf of Riga in Jurmala (Latvia) in March 2017. The algae were identified using a taxonomical identification key (Rudzorga 1995). The samples of algae were washed with deionized water, separated and freeze-dried. Before chemical analyses algae were refined in knife-type mill and then additionally finely ground in a Retsch Mixer Ball Mill MM200.

*Elemental composition and ash content.* The C,H,N, and S contents were measured according to LVS EN 15104:2011 using a Vario MACRO elemental analyzer. Ash content was measured as a residue after treatment at 550 °C in a Carbolite ELF 11/6B furnace.

*Content of extractable compounds.* Sequential extraction with hexane, ethyl acetate (both in Soxhlet apparatus, 8h), boiling 80% aqueous ethanol (batch extraction, 1h x 3 times) followed by hot 2%  $CaCl_2$  solution (70 °C, 1h x 3 times) was performed. The yield of extractable in organic solvent compounds was determined gravimetrically after evaporating the solvent under vacuum. The yield of extractable in  $CaCl_2$  solution compounds was determined by weight difference before and after extraction.  $CaCl_2$  solution after extraction were dialyzed against deionized water (Spectra/Por Dialysis Membrane 6, molecular weight cut-off = 1000 Da) and then freeze-dried.

*Chemical analysis.* The content of phenolic compounds in the extracts was determined by the Folin-Ciocalteu method with gallic acid as reference. Results are expressed in gallic acid equivalent (GAL mg/g of dry extract). Radical scavenging activity was performed against stable radicals DPPH· and ABTS·<sup>+</sup> using spectrophotometric method (Miliauskas *et al.* 2004). Free radical scavenging activity is expressed by IC<sub>50</sub>, which means the concentration required for 50% inhibition of free radicals. Neutral sugar composition was determined using alditol acetate method after treatment with sulfuric acid (Blakeney*et al.* 1983). Alditol acetates were quantified by gas chromatography (Agilent 6850 Series GC System, column TC17) using methyl  $\alpha$ -D-glucopyranoside as an internal standard. The FTIR spectra were recorded with Perkin Elmer Spectrum One spectrometer. LC MS/MS analysis was performed using ACQUITY UPLC H-Class (Waters) with ACQUITY UPLC PDA Detector coupled with SYNAPT G2-Si High Definition Mass Spectrometry (Waters), column – UPLC BEH C18. Mobile phase A: water + 0.1% formic acid, B: acetonitrile, gradient from 5% to 100% B. Mass range (50-1200 Da). EPR spectra were recorded with a Bruker EMX spectrometer operating at 9.6-GHz and a 50-kHz magnetic field modulation.

*The germination test.* The algae and algae containing mixtures (33% algae or solid algae residue after extraction and 67% lignin on o.d.m.) were tested by their effect on cress seeds germination. The seeds were sprouted on a filter paper by wetting it with 50 mg of the tested composition per 10 g of distilled or deionized water. The germination experiments were carried out in three replicates, each with 20 seeds. Cress seeds were sprouted at 20-25 ° C for five days in the dark. The analysis of the sprouted seeds (the length of the shoot and the root) was performed using the WinRHIZO (EPSON PERFECTION 4990 PHOTO scanner).

The biological activity of the algae preparations. The effect of the algae samples under study on growth of the oat seeds was tested. The vegetation tests were carried out in the 50 mL containers, which were filled with the soil substrate "Sulfinor" and placed in the precision climate chamber (Selecta HOTCOLD-GL 2101507). The soil contained > 90% of organic matter, pH 5.5-6.5; Ca -1.1%, Mg 0.06%, humidity 60%, fraction size 0-10 mm. The dose of algae containing samples was  $5-10 \pm 0.1$  g/kg. The oat seeds were placed in the containers filled with the soil at a depth of 1 cm. 20 seeds were used per one experiment. The condition of the germination and growth of the oat seeds were as follows: 18 °C, humidity 90%, luminous intensity 30%, experiment length 15 days. After 15 days a root system analysis (length, average diameter, root volume, count of tips, count of forks) was performed using the WinRHIZO.

#### **RESULTS AND DISCUSSIONS**

Collected seaweeds were identified mainly as *Fucus vesiculosus* – a brown macroalga common on the shores of Baltic Sea. The amount of other algae such as *Furcellaria lumbricalis* was low; therefore they were not analyzed in details in the present study. Elemental composition and ash content of collected *F. vesiculosus* is listed in Table 1 and in general it coincides with literature results (Balina *et al.* 2016).

The results of sequential extraction of *F. vesiculosus* with solvents of increasing polarity have shown that *F. vesiculosus* is low in hexane soluble lipophilic compounds (Table 2). FTIR spectra of bad smelling green hexane extract showed that it contains unsaturated fatty acids (strong absorption at 3010, 2925, 2854, 1740, 1660, 1460, 720 cm<sup>-1</sup> of

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various –CH, -CH<sub>2</sub>, -C=C and ester group vibrations), but the yield of hexane extract was low (~1% o.d.m. of algae). UV and FTIR spectra showed that ethyl acetate and ethanol extract are rich in phenolic compounds which are responsible for free radical scavenging activity. Ethanol extract have a positive reaction with vanillin-HCl typical for *meta*- substituted phenols e.g. in tannins. LC-MS/MS analysis confirmed that ethanol and also ethyl acetate extract contained phlorotannins. Vanillin assay allows estimating tannin content in ethanol extract as approx. 10%.

	Ash. %	C C	ц	N	S	C/N ratio
	ASII, 70	Ľ	п	IN	<u>ہ</u>	C/IN Tatio
F. vesiculosus	$18.5 \pm 0.1$	$39.6 \pm 0.1$	$4.27\pm0.08$	$2.22 \pm 0.01$	$2.30 \pm 0.1$	17.8
<i>F. vesiculosus</i> residue after extraction	$19.7 \pm 0.1$	$38.5\pm0.2$	$4.05 \pm 0.02$	$2.24 \pm 0.03$	$0.63 \pm 0.5$	17.2

Table 1. Ash content and elemental composition of *Fucus vesiculosus* (% of dry weight)

Table 2. Yield of extracts from Fucus vesiculosus, total phenols content in extracts and their antioxidant activity (IC<sub>50</sub>, mg/L).

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Extractant	Yield, % o.d.m.	Phenols, GAE mg/g	DPPH <sup>.</sup> test	ABTS <sup>.+</sup> test		
Hexane	$0.9 \pm 0.2$	n.d.	n.d.	n.d.		
Ethylacetate	$3.1 \pm 0.1$	$430 \pm 30$	28.5	1.8		
80% ethanol	$7.3 \pm 0.4$	$230 \pm 10$	57.7	4.4		
2% CaCl <sub>2</sub>	$10.1 \pm 0.2$	n.d.	> 500 *	52*		

n.d. - not determined, \* - after extract dialysis

Amount of neutral sugars after hydrolysis was higher in more polar ethanol extract (Table 3). Probably sugars present in both extracts as part of glucosides – in ethyl acetate extract mainly as galactosides, but in ethanol extract as mannosides. Ethyl acetate extract contains almost double amount of phenolic compounds than ethanol extract. Radical scavenging activity of ethyl acetate extract against DPPH• and ABTS•<sup>+</sup> radicals is significantly higher than that for ethanol extract. The ABTS•<sup>+</sup> IC<sub>50</sub> value of ethyl acetate extract of *F. vesiculosus* is lower than those for commercial synthetic antioxidant s Trolox, TBHQ, and natural antioxidants, *e.g.*, analogues alder bark extract (Telysheva*et al.* 2011). Thisdemosntrates that *F. vesiculosus* extracts could be potential natural alternative to synthetic antioxidants.

Extraction with CaCl<sub>2</sub> was used to isolate biologically active polysaccharide fucoidan. Obtained fucoidan after dialysis and freeze-drying was pale yellow powder with the yield ~7% from alga. FTIR spectra confirmed presence of sulfate group (absorption at 1230, 845 cm<sup>-1</sup> S=O asymmetric vibration and ester bond) (Wang *et al.* 2010). Elemental analysis showed that fucoidan contained  $7.0 \pm 0.2$  % sulphur or ~20% sulfate groups. Although isolated fucoidan did not exhibit significant antioxidant activity in tests with stable radicals, it is characterized with various biological activities (Ale *et al.* 2011), which made it prospective niche product from *F. vesiculosus*.

Algal biomass contains significant amount of various polysaccharides (Rioux *et al.* 2015). Unlike terrestrial plants, brown algae contain lower amount of neutral sugars. Main monosaccharide of *F. vesiculosus* is fucose. After extraction, total neutral sugar amount increased from  $\sim$ 24% to  $\sim$ 29% (Table 3).

Nitrogen containing compounds content after extraction did not change (Table 1), however relative amount of protein in biomass increased 2.5 times according to the amino acid assay by ninhydrin colorimentric method. Probably part of non-protein nitrogen containing compounds, e.g., chlorophyll, was removed during extraction.

	Fuc*	Ara	Xyl	Man	Gal	Glc	Total
F. vesiculosus	$10.9 \pm 1.0$	$0.5 \pm 0.1$	$2.0 \pm 0.1$	$2.8 \pm 0.4$	$1.8 \pm 0.1$	$6.6 \pm 0.3$	24.6
<i>F. vesiculosus</i> <i>e</i> thyl acetate extract	$0.47 \pm 0.09$	0.36 ± 0.19	-	$0.46 \pm 0.05$	$2.74 \pm 0.16$	-	4.1
<i>F. vesiculosus</i> ethanol extract	$0.18\pm0.07$	$0.44 \pm 0.14$	-	8.7 ± 0.5	$1.11 \pm 0.05$	$0.84\pm0.04$	11.3
<i>F. vesiculosus</i> after extraction	$12.1 \pm 1.2$	$0.5 \pm 0.1$	$2.7 \pm 0.2$	$3.0 \pm 0.4$	$1.9 \pm 0.2$	8.7 ± 0.4	28.9

Table 3. Neutral sugar composition of Fucus vesiculosus before and after extraction and in extracts (% of dry weight).

\* Fuc - fucans, Ara - arabinans, Xyl - xylans, Man - mannans, Gal - galactan, Glc - glucans.

Relatively low C/N ratio detected for *F. vesiculosus* biomass did not change significantly during extraction and was comparable with commercial grass compost (Bikovens *et al*, 2010). The addition of algae to tested mixture showed beneficial effect on cress (*Lepidium sativum*) seeds germination and root system development. The addition of the algal biomass to soil promoted the growth and development of the root system of the oat (*Avena sativa*) seeds. The effect of *F. vesiculosus* was dose-depended and was noticeable after 5 g/kg addition. Oat root length, amount of roots tips and forks increase were more prominent when *F. vesiculosus* residue after extraction was used (Table 4). The comparison of the unextracted algae and algae residue, obtained after extraction of fucoidan, showed, that the latter has stronger positive effect on the oat seeds growth and germination.

As alternative approach for processing of the *F. vesiculosus* biomass, the mechanochemical treatment of algae biomass with hydrolyses lignin was proposed. EPR analyses confirmed interaction between algae biomass and lignin. Free spin concentration in algal-lignin complex is more than 2 times lower  $(4.3*10^{15} \text{ spin/g})$  in comparison with *F. vesiculosus* biomass  $(1.2*10^{16} \text{ spin/g})$  and lignin  $(1.1*10^{16} \text{ spin/g})$ . Currently the vegetation tests are carried out for estimation of novel fertilizer effects on plants growth and development.

	Root length, cm	Average diameter,	Root volume, cm <sup>3</sup>	Count of tips	Count of forks
		mm			
Control	$55 \pm 9$	$0.40 \pm 0.05$	$0.10 \pm 0.02$	$394 \pm 54$	$294\pm71$
<i>F. vesiculosus</i> in the soil 5 g/kg	61 ± 12	$0.40\pm0.05$	$0.10\pm0.02$	$372 \pm 48$	$345\pm 64$
<i>F. vesiculosus</i> in the soil (10 g/kg)	$88 \pm 14$	$0.40\pm0.05$	$0.10 \pm 0.02$	$688 \pm 61$	$338\pm52$
<i>F. vesiculosus</i> after extraction in the soil (5g /kg)	$70 \pm 7$	$0.40 \pm 0.04$	$0.10 \pm 0.02$	$664 \pm 56$	$353 \pm 42$

Table 4. Effect of addition of Fucus vesiculosus in the soil on oat growth.

## CONCLUSIONS

The present work proposes complex processing of the brown algae *Fucus vesiculosus* biomass, which can be collected at the costs of the Baltic Sea. The promising approach includes the extraction of the fucoidan, phenolic compounds with biological and antioxidant activities important for pharmaceutical, cosmetic and food industries and the use of the residue after extraction as a fertilizer in agriculture. The studied algae biomass is not phytotoxic, promotes the growth of the agricultural plants and has a good potential in sustainable agriculture. The mechanochemical treatment was proposed for obtaining novel fertilizer based on algal biomass.

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